General Microbiology Laboratory (PMB112L), Spring '07

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Lab: Monday, 1:00 - 5:00 PM, Room 209 Genetics and Plant Biology Bldg.

Discussion: Wednesday, 5:00 - 6:00 PM, Room 2319 Tolman Hall.

Content: Experimental techniques of microbiology designed to accompany the lectures in C112 and C148. The primary emphasis in the laboratory will be on the cultivation and physiological and genetic characterization of bacteria. Laboratory exercises will also include the microscopic observation and staining of bacteria from selected environments.

We will learn many principles with a laboratory isolate of the intestinal bacterium *E. coli*. Some reasons for this are listed below. Our environmental exercises will focus on microorganisms found in marine sediment.

We will emphasize *E. coli* for the following reasons:

1. *E. coli* is arguably the best-studied and best understood cell on the planet. Because I have years of experience with *E. coli* and other enteric bacteria it is easiest for me to teach you simple principles of working with a laboratory organism by using *E. coli* K12.

2. There is a complete genome sequence for *E. coli* and my laboratory studied expression of all of its genes simultaneously using DNA microarrays. We will analyze a DNA microarray experiment performed in my laboratory.

3. What do you think is not known about *E. coli*? What is known? Why would a serious biologist continue to work with laboratory strains of *E. coli*? (See the attached article by H. Mori.)

4. Non-laboratory strains of *E. coli* can be pathogenic. As you know, some (e.g. *E. coli* O157H7) can cause extremely serious intestinal disease. Other strains, which are thought to be intestinal commensal strains of humans, are a prominent cause of urinary tract infections. Such infections occur frequently in women.

5. As a robust intestinal organism, *E. coli* is used as an indicator of fecal contamination in drinking water and during purification of waste water. A number of fecal contaminants that are transmitted by the so-called oral-fecal route cause serious intestinal disease. When *E. coli* is found in drinking water, one must worry about the presence of pathogenic fecal contaminants generally.

6. The *E. coli* genome has been pared down in organisms of the genus Buchnera, which are obligate intracellular symbionts of aphids. The Buchnera genome contains ~500 genes, as opposed to the ~4400 of *E. coli*. These genes represent alternative minimal gene sets required for life. (See attached abstracts.)

Readings: I will give you appropriate readings from the text for C112/C148 [Brock Biology of Microorganisms, Tenth edition, M. T. Madigan, J. M. Martinko, and J. Parker. Prentice Hall, Upper Saddle River, NJ, 2003 = "Brock"] and from two other texts [Microbiology, a Human Perspective, E. W. Nester, C. E. Roberts, M. T. Nester. Wm. C. Brown, Dubuque, Iowa, 1995 = "Nester"] [Biology of the Prokaryotes, J. W. Lengeler, G. Drews, and H. G. Schlegel, editors. Blackwell Science, New York, 1999 = "Schlegel"], as useful.

Grades, Preparation, Discussion Sections: Grades will be based on laboratory reports and performance in the laboratory. It is essential that you read the exercises and prepare protocols before the laboratory begins. Be prepared to ask questions on aspects of the laboratory that are confusing or that require amplification. **Wednesday evening discussion sections are REQUIRED.**

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Review

From the Sequence to Cell Modeling: Comprehensive Functional Genomics in *Escherichia coli*

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As a result of the enormous amount of information that has been collected with *E. coli* over the past half century (e.g. genome sequence, mutant phenotypes, metabolic and regulatory networks, etc.), we now have detailed knowledge about gene regulation, protein activity, several hundred enzyme reactions, metabolic pathways, macromolecular machines, and regulatory interactions for this model organism. However, understanding how all these processes interact to form a living cell will require further characterization, quantification, data integration, and mathematical modeling, systems biology. No organism can rival *E. coli* with respect to the amount of available basic information and experimental tractability for the technologies needed for this undertaking. A focused, systematic effort to understand the *E. coli* cell will accelerate the development of new post-genomic technologies, including both experimental and computational tools. It will also lead to new technologies that will be applicable to other organisms, from microbes to plants, animals, and humans. *E. coli* is not only the best studied free-living model organism, but is also an extensively used microbe for industrial applications, especially for the production of small molecules of interest. It is an excellent representative of Gram-negative commensal bacteria. *E. coli* may represent a perfect model organism for systems biology that is aimed at elucidating both its free-living and commensal life-styles, which should open the door to whole-cell modeling and simulation.

Keywords: *E. coli*, Genome, Modeling, Systems biology

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The *E. coli* Genome Sequence

The complete *E. coli* genome sequence was determined through independent efforts by American and Japanese groups using two different strains of *E. coli* K-12, MG1655 and W3110 (Aiba *et al.*, 1996; Itoh *et al.*, 1996; Oshima *et al.*, 1996; Blattner *et al.*, 1997; Yamamoto *et al.*, 1997). These strains were diverged from the same ancestral strain about 50 years ago, resulting in slight but significant differences, including the large inversion involving the ribosomal RNA genes (see below). A complete genome sequence analysis revealed precise differences between the two strains. A comparison of the genome sequence has revealed relatively infrequent, 1 per 105 base substitution mutations in the protein coding region (Itoh *et al.*, 1999). Recently, a Japanese group confirmed the differences between the genomes of MG1655 and W3110 by PCR-based direct sequencing. They found that only 9 bases in 8 different ORFs represent actual sequence differences besides large scale insertions or deletions, such as IS elements (Horiuchi, in preparation). In summary, the *E. coli* genome seems to tolerate large rearrangements such as insertion, deletion and recombination better than micro-scale changes like base substitution. In 2001, the pathogenic *E. coli* strain O157, which is very closely related to *E. coli* K-12, was subjected to genomic sequencing by two groups (Hayashi *et al.*, 2001; Perna *et al.*, 2001). The results obtained with this pathogenic strain agreed well with the above findings with strain K-12. *E. coli* O157 appears to have acquired its pathogenicity mainly by horizontal gene transfer that is mediated by a temperate bacteriophage.

Prediction of ORFs from the complete genome sequence of *E. coli* While more than 400 bacterial genomes have been sequenced, the functional assignment of all the gene products has not yet been accomplished for any microorganism. Extensive studies in a few model organisms, including

Escherichia coli and *Saccharomyces cerevisiae*, elucidated the functions of many genes and gene products, particularly by using techniques of genetics, biochemistry and physiology, etc. *E. coli* K12, the best-studied microorganism, is estimated to contain about 4,400 genes of which about 2,000 have not been characterized experimentally (Mori *et al.*, 2000). Altogether about 3,700 genes can be assigned or predicted a function with reasonable assurance, based on biochemical experiments and computational analysis in *E. coli* and other microorganisms. Of the remaining approximate 700 genes, 650 show sequence similarity to genes of unknown function in other bacteria, whereas 50 show no obvious similarity to any known genes. The assignment of function to these unknown genes is one of the major targets of functional genomics in *E. coli*. In addition to their fundamental importance for understanding *E. coli* biology, these functional assignments are significant for three other reasons: (1) complete functional assignment will result in the discovery of new physiological and biochemical pathways, (2) will facilitate functional assignment in other bacteria, and (3) will lead to the identification of new targets for antibiotic design other than for biotechnological development.

Besides, elucidation of individual gene functions, which is basic to the understanding of a cell, systematic analysis of relationships between genes or gene products is also a significant target that is just starting to be explored systematically.

The availability of numerous complete genome sequences has considerably accelerated the comparative approach. Various analyses for clustering genes (ORFs) have been performed. New concepts have been proposed such as ancient conserved regions (ACR) (Koonin *et al.*, 1995), clusters of orthologous groups (COGs) (Tatusov *et al.*, 1997) and modules (Riley and Labedan, 1997). These analyses are quite valuable, not only from an evolutionary point of view, but also from a more practical view point as well, such as the functional prediction of hitherto uncharacterized ORFs.

A workshop on the Annotation of *Escherichia coli* K-12 2003, chaired by Monica Riley, was recently held at Woods Hole, MA, on November 14-18. Fourteen scientists from the US, Europe and Japan gathered and took parts in two tasks. One group addressed the annotation of known and predicted gene products. The other group focused on gene boundaries and sequences. The intensively curated and coordinated data will be submitted to GenBank in early 2004, and made available on the Internet for public access (M. Riley, personal communication).

Repetitive sequences, sites, RNA genes etc. Many kinds of repetitive sequences are found within the genome, some of which have important physiological functions. The distribution of repeats along the chromosome is not random and seems to be related to some feature of DNA replication. Repetitive sequences in the *E. coli* genome are encountered in different contexts. Various classes of repeats are present in diverse prokaryotes, including *E. coli*. Coding sequences such as ribosomal RNA genes, transfer RNA genes, and insertion sequences are usually present in multiple copies, but their copy numbers are relatively low. Other interspersed repetitive DNA sequences are relatively short but abundant and located within intercistronic non-coding sequences. *E. coli* has 7 copies of rRNA coding genes (*rrn*) and an additional copy of the 5S rRNA gene (Blattner *et al*., 1997). Other rRNA-related DNA sequences (TRIP) showing significant similarity to 5S rRNA were recently discovered. These may have important functions that are related to 5S rRNAs (Rudd, 1999). The rRNA genes are located within half of the chromosome that contains the origin of DNA replication (*oriC*), whereas many of the TRIP sequences are located in the other half of the chromosome. Moreover, *rrn* and some of the related TRIP sequences are distributed symmetrically on the leading strands on both sides of *oriC*. Consistent with this location, the transcription of *rrn* operons generally proceeds away from the replication origin. Repetitive sequences, such as rRNA genes, provide a driving force for genome rearrangement. Large inversions of the genome between rRNA genes (between *rrnD* and *rrnE*) have been documented in *E. coli*, and one such inversion was found in the W3110 strain (Hill and Harnish, 1981). Although the direction of *rrn* gene transcription is strictly oriented away from the replication origin, the inversion in *E. coli* appears to be stable because the geometric relationship between *rrn* operons and the replication origin are preserved. Insertion Sequences (IS) represent another family of repetitive elements and cause genetic variation among different strains of *E. coli*: both the abundance and distribution of insertion sequences can vary in different strains (Mahillon and Chandler, 1998; Mahillon *et al.*, 1999). The W3110 strain contains about 60 ISs that belong to at least 10 distinct families. At least 10 of these ISs differ in their location when compared to MG1655 strain. These results testify to the great variability in both number and family of ISs in closely related strains. Interspersed repetitive sequences represent relatively short (usually less than 500 bp), non-coding, intercistronic and dispersed elements that are found in bacterial genomes. Six classes of highly repetitive sequences, BIME, IRU (Sharples and Lloyd, 1990), Box C (Bergler *et al.*, 1992), RSA (Mizobuchi, personal communication), iap (Ishino *et al.*, 1987; Nakata *et al.*, 1989), and Ter Sequences (Hill, 1996) have been identified (Table 1). These sequences were

Table 1. Extragenic highly repetitive sequence families

Sequence	Size (bp)	Copy number
BIME	40~400	~800
Ter	30	63
BoxC	56	36
Iap	29	23
IRU	127	19
RSA	152	6

primarily discovered by computer analysis of sequence data. None of these sequences encode proteins. They are dispersed throughout the chromosome.

Finally, a number of RNAs that do not function as either mRNAs, tRNAs, or rRNAs have been discovered, mostly fortuitously. The non-mRNAs have been predominantly termed as small RNAs in bacteria. The potentially important function of some of them was recently documented (Storz, 2002). Systematic analysis of non-coding RNA in *E. coli* has just started (G. Storz, personal communication).

Post Genome Sequencing Project

As previously described, the function of nearly half of the total ORFs in *E. coli* is not unknown, of which 20% remain difficult even to predict their function. The latest estimates reveal that about 700 to 800 of total ORFs have no attributable function. Therefore, a high priority will be placed on the development of novel, high-throughput technologies to identify their function. A new comprehensive "molecular tool kit" would be required to define unknown gene functions and to assign potentially new roles to known genes. A large number of valuable plasmid constructs, *E. coli* strains, assay tools, and other biological materials that are useful for analyzing the gene function have been constructed, developed, collected, and tested. These will form key resources for getting basic and comprehensive knowledge on *E. coli* biology and for exploiting these data with *E. coli* for other organisms. Furthermore, methods for genome-wide analysis of transcriptional regulation (transcriptome), protein dynamics (proteome) and flow of metabolites (metabolome) have rapidly evolved to make the best use of the rich DNA sequence information ("-ome" is a Greek suffix for "whole").

Experimental resources A DNA sequence analysis has identified about 4,400 protein-coding genes in *E. coli*. The power of genetically- tractable model organisms resides in the fact that they can facilitate the global and systematic analysis of physiological gene function *in vivo*. Precise genetic manipulation is particularly important for functional genomics. Genome sequence data has permitted the design of oligo DNA primers for the precise amplification of entire ORFs and generation of a complete set of histidine-tagged ORF clones (with or without fusion to the GFP gene) (Mori *et al.*, 2000).

One approach for systematic functional analysis is to make use of gene deletion (replacement) that is obtained by homologous recombination. Targeted gene replacement, once thought to be difficult in *E. coli*, can now be dealt with by using the techniques that were developed by Wanner and colleagues (Datsenko and Wanner, 2000). The systematic attempts to construct these resources for functional genomics is now rapidly raising *E. coli* to one of the leading organisms in the field of functional genomics.

ORF clones

ORF clones will provide a basic genetic tool for studying gene function, since they provide a template for PCR amplification and for preparing purified gene products etc. To clone all of the genes of *E. coli*, a plasmid vector with the following properties was constructed: (1) high copy number plasmid, (2) IPTG inducible expression of cloned ORF and repression of expression by *lac*I^q, (3) a Histidine tag coding sequence attached to the N-terminal of ORF, (4) in-frame fusion with GFP coding sequence at the C-terminal end, (5) generation of *Sfi*I restriction sites at both boundaries of the cloned ORF, (6) possibility of GFP fragment removal by *Not*I (Kitagawa, in preparation). The whole set of PCR amplified ORF fragments were cloned into the *Stu*I site of this vector. As far as we know, these clones represent the only comprehensive collection of *E. coli* ORFs that is currently publicly available. Fig. 1 shows the structure of these clones.

Knock-out mutants

A systematic mutational analysis of genes in their chromosomal location should provide basic information and insight into their function. A large number of these mutants have already been established, primarily by random insertion mutagenesis (Miki *et al.*, personal communication). Although this effort provided the research community with a unique collection of mutants, it was necessary to determine the insertion position of the transposons. In addition, some complications could not be avoided due to the nature of transposon mutagenesis, such as incomplete disruption of the targeted gene or polar effects on the downstream genes. Regarding this last point, only the set of in-frame deletion mutants can avoid this problem. Most bacteria, including *E. coli*, are not readily transformable with linear DNA because of the presence of intracellular exonucleases that degrade transformed linear DNA. In contrast, genes can be directly disrupted in *Saccharomyces cerevisiae* by transformation with PCR fragments encoding a selectable marker having only 35 nt of flanking DNA homologous to the chromosome (Baudin *et al.*, 1993). On the other hand, it has long been known that many bacteriophages encode their own homologous recombination systems (Smith, 1988). It was recently shown that the λ Red (*g, b, exo*) function promotes a greatly enhanced rate of recombination over that exhibited by the *recBC sbcB* or *recD* mutants when using linear DNA. Wanner and his colleagues developed a convenient procedure based on the l Red system that provides an efficient way to isolate replacement mutants using PCR fragments encoding an antibiotic resistance gene and having only 40 to 50 nt of flanking regions (Datsenko and Wanner, 2000). A comprehensive and clean deletion mutant library for all *E. coli* ORFs (the KO collection, Knock-Out and KEIO University) using the λ Red system is now under construction and will be open to the public soon (Baba, in preparation). The design of isolating these deletions is illustrated in Fig. 2.

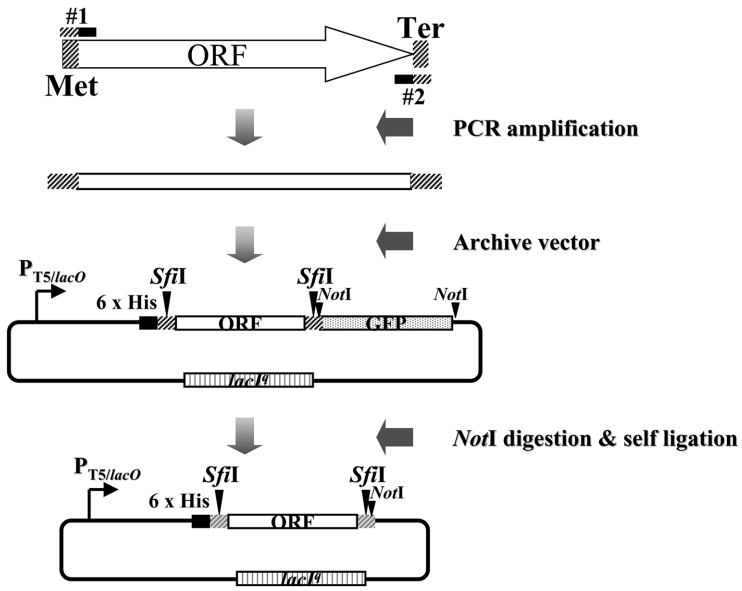

Fig. 1. Construction of Archive clones.

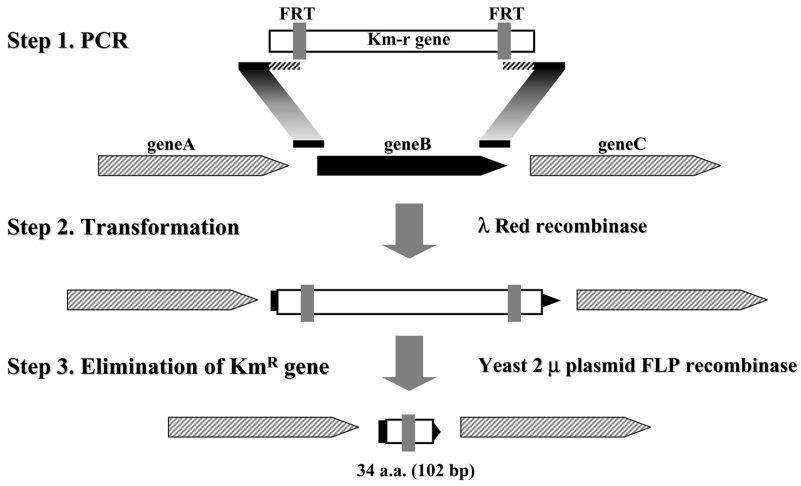

Fig. 2. Construction of deletion mutants.

Large deletion mutants

Two recent publications describe deletions of large chromosomal segments from the *E. coli* genome. Kim and his colleagues made two large libraries of independent transposon mutants using modified Tn5 transposons with two different selection markers, thereby precisely mapping their chromosomal location (Yu *et al.*, 2002). This method allows the integration of the mapped transposon insertion carrying *loxP* site from each of the mutant libraries into the same chromosome followed by excision of the flanking genomic segments by site-specific Cre mediated *loxP* recombination. Alternately, Blattner and his colleagues used a deletion "by design approach" and have already generated lineages in which more than 10% of the genome has been eliminated without loss of viability (Kolisnychenko *et al.*, 2002). Another large-scale deletion construction has been undertaken by Katoh (J. Katoh, personal communication). Basically, the approach consists of markerless gene replacement, and has already generated a genome lacking more than 25% of the original genome DNA. Intermediates have been saved and descendents with larger deletions are being generated. By examining representatives of these collections for growth defects under specific environmental conditions, the effects of losing many genes can be traced simultaneously. Data generated in this type of experiment will be a great asset for cell modeling. These approaches are likely to be especially advantageous when single gene mutations display no discernable phenotypic changes.

Other approaches for functional geneomics

Two different types of technology were recently developed to study protein expression and protein-protein interaction. One

is based on novel tandem affinity purification (TAP) of tag fusion protein. This is a generic procedure to purify target proteins expressed at their natural level under native conditions (Rigaut *et al.*, 1999). To investigate heteromeric protein complexes of unknown composition, standard systems for protein overexpression may lead to the assembly of overexpressed proteins as non-physiological complexes. To overcome this problem, a TAP tag fusion cassette was developed which encodes the calmodulin-binding peptide (CBP), a TEV protease recognition site, and proteinA of *Staphylococcus aureus* (ProtA).

The other method depends on the genome-wide, registered collection of *E. coli* bioluminescent reporter gene fusions. Each of the random fusions of *E. coli* chromosomal DNA fragment to the *Photorhabdus luminescens luxCDBE* reporter gene was precisely mapped by sequencing (Van Dyk *et al.*, 2001). To identify and quantify changes in the expression level, the authors tested this type of fusion and analyzed alterations in the expression levels of heat shock, SOS response and oxidative stress genes.

Transcriptome analysis To exploit the rapid progress in genome research, many novel techniques have been developed including DNA microarray or DNA chip technology that are extremely useful for the analysis of global gene expression (Lockhart *et al.*, 1996). Generally, a DNA microarray is defined as an orderly arrangement of tens to hundreds of thousands of unique DNA molecules of known sequence, usually on a glass slide. Unique DNA molecules are either individually synthesized on a rigid silicon plate (generally referred to as DNA chips and developed by Affymetrix Co.) or prepared from pre-synthesized DNA (synthetic oligo-nucleotides or PCR products) that are spotted and immobilized on a slide glass. The use of DNA microarrays to study *E. coli* gene regulation was first illustrated by Blattner and colleagues. This has rapidly expanded and been applied to study various aspects of transcriptional regulation (Tao *et al.*, 1999). To elucidate the whole transcriptional regulatory network, several systematic approaches using DNA microarray have been performed (Oshima *et al.*, 2002; Masuda and Church, 2003). These analyses were done not only under different growth conditions but also using either the overexpression or deletion of a target regulatory gene. As previously described, large experimental resources including complete sets of clones and deletion mutants of all of the predicted genes of *E. coli* are being established. These resources will contribute in the acceleration of the systematic transcriptome analyses. Accumulation of the results from such these large-scale analyses using DNA microarrays will also assist more traditional biological research, as well as the construction of large databases that will be very beneficial. Public availability of this data is not limited to supplemental data derived from publications on the ftp site of journal publishers, but is also available from integrated databases such as the KEGG database as a systematic collection of microarray data (Kanehisa *et al.*, 2002).

The accumulation of publicly-available DNA microarray analyses data promotes rapid computational analysis of gene expression profiles. In general, bacterial genes form operon in which multiple ORFs are transcribed from the same promoter to form a single mRNA transcript. Prediction of operons using DNA microarray experiments is one approach used to reconstruct gene regulatory networks at the whole genome scale (Sabatti *et al.*, 2002). Reconstruction of global regulatory networks using genome-scale gene expression data sets has also been reported recently (Gutierrez-Rios *et al.*, 2003; Herrgard *et al.*, 2003).

Proteome One of the basic technologies for global analysis of cellular proteins has been developed by O'Farrell as two-dimensional polyacrylamide gel electrophoresis (O'Farrell, 1975). *E. coli* has a long history of protein cataloging by 2D gel (Vanbogelen, 1996). The major limitations of 2D electrophoresis are related to the difficulty in assigning gene identities to observed spots and the lack of spots for proteins that do not separate well on 2D gels. In addition, the method displays a general bias against membrane proteins and proteins of low abundance. However, matrix-assisted laser desorption ionization-mass spectrometry (MALDI-MS) has greatly contributed to the acceleration of protein identification (Figeys *et al.*, 1998). A new gel based separation system (RFHR) has also significantly expanded the separation range of proteins (Wada *et al.*, 1993). The assignment of 2D gel data will be coordinated between two major ongoing initiatives, the CyberCell project (Ellison, personal communication) and the efforts of a Japanese group (Wada, personal communication). iIn addition, several alternative methods, based on an analysis of peptides from whole cell lysates by LC/MS, were developed and successfully used (Gygi *et al.*, 2000; Corbin *et al.*, 2003).

Protein localization

Even though the bacterial cell have no complex intracellular compartments that are hallmarks of eukaryotic cells, there is comprehensive information about the location of proteins within the bacterial cell, which is important for understanding their functions and interactions. Large-scale analyses of protein localization in *S. cerevisiae* have been reported (Ross-Macdonald *et al.*, 1999; Kumar *et al.*, 2002). Recently, Niki *et al.* also reported the comprehensive analysis of protein localization using clones of individual ORFs that were fused with GFP protein under non-induced growth conditions (Niki, personal communication). Localized GFP fluorescence was successfully observed for about 4,000 out of 4,300 genes tested. The patterns of localization were classified roughly into 4 distinct categories. Protein localization, based on subcellular fractionation, is also underway as part of the CyberCell project (Ellison, personal communication) and in parallel at Harvard University (G. Church, personal communication).

Protein-protein interaction
Most cellular processes are carried out by multiprotein complexes. The identification and analysis of these protein complexes provide further insight into the physiological function and molecular mechanisms of the functional units. Following identification and cataloging of all of the proteins that are expressed in a cell, a global analysis of the protein-protein interaction becomes critical for understanding cellular processes. In *S. cerevisiae*, a genome-scale analysis of the protein complexes was performed using the yeast two-hybrid system (Fromont-Racine *et al*., 1997; Uetz *et al.*, 2000; Ito *et al.*, 2001), protein chips (Zhu *et al.*, 2001), or affinity tagged system (Gavin *et al*., 2002). In *E. coli*, there are presently two comprehensive analyses underway using affinity-tagged proteins, chromosomally tagged with TAP (J. Greenblatt, personal communication) and plasmid clones containing an histidine tag (Arifuzzaman, in preparation), to identify the protein-protein interaction, as was done in *S. cerevisiae*. These will allow identification by mass spectrometry of proteins that co-purify with the tagged baits that are thus candidate-interacting proteins. Greenblatt and his colleagues are focusing on studying the interaction of nearly 200 highly conserved proteins that are known to be essential, such as DNA and RNA polymerases. They have developed an interaction network for these proteins. On the other hand, Arifuzzaman and his colleagues performed high-throughput analysis using plasmid clones, although most of the membrane proteins were only poorly purified and failed to function as bait. However, out of 4,300 total ORFs, more than 2,700 ORFs were successfully purified from the plasmid clones and candidates that can interact with His-tagged bait proteins were identified. The total number of observed interactions in this set of 2,700 proteins amounts to an impressive total of about 14,000 potential interactions (Arifuzzaman, in preparation). An analysis of this complex protein interaction network is now underway.

Metabolome A substantial portion of the *E. coli* genome encodes enzymes that interconvert metabolites, synthesize cofactors, and regulate small molecule metabolism. Metabolites can in turn control the gene expression and are allosteric regulators of enzymes. The metabolome can be described as the total complement of metabolites in a cell (Tweeddale *et al.*, 1998). Metabolome analyses can be performed using several approaches, such as metabolite profiling, flux analysis using isotopic tracer, and pathway reconstruction etc. This allows insight into metabolic and physiological responses within a cell. Global metabolite profiling will provide deeper insight not only into metabolism but also into cellular physiology and functional genomics (Fiehn, 2002). This approach has been used for functional genomics studies in plants (Fiehn *et al.*, 2000) and yeast (Raamsdonk *et al.*, 2001). In *E. coli*, metabolites were labeled with C-14 glucose and identified by 2-dimensional thin-layer chromatography after extraction by cold methanol (Maharjan and Ferenci, 2003). Using this approach, the authors identified about 100 metabolite spots. Recently, Soga and colleagues developed a powerful analytical method using capillary electrophoresis-electrospray ionization mass spectrometry (CE-ESI-MS) that dramatically increases the number of metabolites that can be measured simultaneously (Soga *et al.*, 2002a, 2002b, 2003).

E. coli Genome as a Model for Systems Biology

Biology itself is now at a turning point between the past descriptive science and the emerging modern quantitative systems biology. Understanding the causal relationships between the genotype and phenotype will require a very significant expansion of the traditional toolbox that is used by molecular biologists. It must include concepts and techniques from many other scientific disciplines such as physics, mathematics, numerical analysis, stochastic processes, and control theory. Many novel tools must be developed to understand how dynamic, robust but adapting, and developing systems can emerge from the information buried in the genome (Ehrenberg *et al.*, 2003). The genome sequencing efforts and the subsequent bioinformatics analyses have not only defined the molecular parts for a number of living organisms, but also opened up possibilities to reconstruct the metabolic pathways. The stoichiometric coefficients for each enzyme in the *E. coli* metabolic map were assembled to construct a genome-specific stoichiometric matrix. This matrix was used to define the systems characteristics and the capabilities of this organisms metabolism (Edwards and Palsson, 2000). In that report, the authors showed the result of comparisons between the in silico predictions and experimental observations using deletions of genes in the central metabolic pathways. These approaches have now been expanded to the genome-scale reconstruction not only of metabolic network (Forster *et al.*, 2003; Reed *et al.*, 2003), but also of heterogeneous network types including transcription and translation (Shen-Orr *et al.*, 2002; Allen *et al.*, 2003; Gutierrez-Rios *et al.*, 2003). A shift in biology from a component-based perspective to a systems view of the cell is occurring based on genome sequence accumulation and high-throughput post-genomic data generation. Modeling cellular functions according to a systems biology is not new but this approach is now expanding to reach the genome-scale. The total number of genes and biochemical elements that are integrated into a single model has now reached ~2000 (Reed and Palsson, 2003). In parallel, several software environments for the quantitative simulation of cellular processes, including metabolic pathways, based on the numerical integration of rate equations, have been developed (Goryanin *et al.*, 1999; Tomita *et al.*, 1999; Mendes and Kell, 2001; Hucka *et al.*, 2003).

Table 2. Useful websites for *Escherichia coli*

http://ecoli.aist-nara.ac.jp	GenoBase database at Nara Institute of Science and Technology, Japan
http://www.genome.ad.jp	KEGG database at Kyoto University, Japan
http://www.shigen.nig.ac.jp/ecoli/pec/index.jsp	PEC database at the Institute of National Genetics, Japan
http://gib.genes.nig.ac.jp	GIB database at the Institute of National Genetics, Japan
http://genome.gen-info.osaka-u.ac.jp/bacteria/o157	Web site of *E. coli* O157 at Osaka University, Japan
http://www.cifn.unam.mx/Computational_Genomics/regulondb	RegulonDB at Universidad Nacional Autonoma de Mexico, Mexico
http://redpoll.pharmacy.ualberta.ca/CCDB	CyberCell Project at University of Alberta, Canada
http://www.uni-giessen.de/~gx1052/ECDC/ecdc.htm	*E. coli* Database Collection at Justus-Liebig-University, Germany
http://genolist.pasteur.fr/Colibri	Colibri database at the Institute Pasteur, France
http://web.bham.ac.uk/bcm4ght6/res.html	The *E. coli* Index at the University of Birmingham, UK
http://colibase.bham.ac.uk	Colibase dataset at the University of Birmingham, UK
http://us.expasy.org/ch2d	SWISS-2DPAGE in SWISS-PROT database at Swiss Institute of Bioinformatics
http://kr.expasy.org/enzyme	Enzyme database at Swiss Institute of Bioinformatics
http://www.EcoliCommunity.org	*E. coli* Community at Purdue University, USA
http://biocyc.org/ecocyc	EcoCyc database at SRI International, USA
http://www.genome.wisc.edu	Genome Project at University of Wisconsin ñ Madison, USA
http://bmb.med.miami.edu/EcoGene/EcoWeb	EcoGene database at University of Miami School of Medicine, USA
http://www.vetsci.psu.edu/ecoli.cfm	*E. coli* Reference Center at Penn State University, USA
http://www.ncbi.nlm.nih.gov	National Center for Biotechnology Information, USA
http://cgsc.biology.yale.edu	*E. coli* Genetic Stock Center at Yale University, USA
http://www.ecoli.princeton.edu/index.php	*E. coli* Bioinformatics/Resources Initiative at Princeton University, USA
http://genomics.lbl.gov/~ecoreg/index.html	EcoReg, The *Escherichia coli* Regulation Consortium, USA

International Consortium for Large Scale *E. coli* Modeling

The International *E. coli* Alliance (IECA, http://www.EcoliCommunity.org) was formed in November 2002 to tackle the fundamental biological problem in developing the first comprehensive computational model of a living cell. IECAs mission is to consolidate global efforts to understand a living bacterial cell. Scientists around the world are working together to create a complex computer model, integrating all of the dynamic molecular interactions that are required for the life of a simple, self-replicating cell. An *E. coli* cell model will have immediate practical benefits in biology and bioengineering and should significantly contribute to advancing the field of computational systems biology. The generation of a computerized *E. coli* cell will also add powerful new tools to our existing arsenal for functional discovery, including virtual experimentation and mathematical simulation. Ultimately, these biological and computational tools could be useful in both drug discovery and in the design of bioenhanced nanomachines. Furthermore, the development of a virtual system for experimentation on the *E. coli* cell will be extremely useful for understanding more complex cells and contribute to the development and validation of in silico models of human cells and whole multicellular organisms. Biology is now evolving to become a “big science”. The tiny *E. coli* is well positioned to become one of the giant players in the new biology era, based on the determining role it played in the field of molecular genetics.

Useful *E. coli* Websites

As internet technology advances, and the scale of experimental approaches grows exponentially, the importance of biological information and data repository websites cannot be overstated. Some useful websites for *E. coli* biology are listed in Table 2.

Epilog

Robert Hooke first used the term “cell” to describe the basic structural unit of cork in 1665. Biology still has a long way to go for a complete understanding of a cell, even though the complete genetic blueprints are available. In the last four centuries, biology has developed and supported intensive activities based on traditional small-scale research. This type of research will always be important and needs to increase in the future in order to build a more precise quantitative model of biological processes or a cell itself. On the other hand, it is also absolutely true that solving large scale and complex biological networks is far beyond the conventional approach. As previously, biology itself is now standing at a turning point. I hope these new approaches will flower with traditional ones, because these are exactly complementary.

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FULL FINAL TEXT OXFORD JOURNALS

Phloem-sap feeding by animals: problems and solutions.

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The incidence of phloem sap feeding by animals appears paradoxical. Although phloem sap is nutrient-rich compared with many other plant products and generally lacking in toxins and feeding deterrents, it is consumed as the dominant or sole diet by a very restricted range of animals, exclusively insects of the order Hemiptera. These insects display two sets of adaptations. First, linked to the high ratio of non-essential:essential amino acids in phloem sap, these insects contain symbiotic micro-organisms which provide them with essential amino acids. For example, bacteria of the genus Buchnera contribute up to 90% of the essential amino acids required by the pea aphid Acyrthosiphon pisum feeding on Vicia faba. Second, the insect tolerance of the very high sugar content and osmotic pressure of phloem sap is promoted by their possession in the gut of sucrase-transglucosidase activity, which transforms excess ingested sugar into long-chain oligosaccharides voided via honeydew. Various other animals consume phloem sap by proxy, through feeding on the honeydew of phloem-feeding hemipterans. Honeydew is physiologically less extreme than phloem sap, with a higher essential:non-essential amino acid ratio and lower osmotic pressure. Even so, ant species strongly dependent on honeydew as food may benefit from nutrients derived from their symbiotic bacteria Blochmannia.

Publication Types:

- Review

PMID: 16449374 [PubMed - indexed for MEDLINE]

2: Heredity. 2005 Nov;95(5):339-47. Related Articles, Links

Chromosomal stasis versus plasmid plasticity in aphid endosymbiont Buchnera aphidicola.

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The study of three genomes of the aphid endosymbiont Buchnera aphidicola has revealed an extraordinary stasis: conservation of gene order and genetic composition of the chromosome, while the chromosome size and number of genes has reduced. The reduction in genome size appears to be ongoing since some lineages we now know to have even smaller chromosomes than the first B. aphidicola analysed. The current sequencing by our group of one of these smaller genomes with an estimated size of 450 kb, and its comparison with the other three available genomes provide insights into the nature of processes involved in shrinkage. We discuss whether B. aphidicola might be driven to extinction and be replaced by secondary aphid endosymbionts. In some lineages, genes encoding key enzymes in the pathways leading to tryptophan and leucine biosynthesis (trpEG and leuABCD, respectively) are located on plasmids, rather than the main chromosome. In contrast to the stasis of the main chromosome, plasmid genes have frequently been transferred to the main chromosome and undergone other gene rearrangements. We propose a two-step scenario to explain these contrasting modes of evolution. Essential genes may have escaped regulation by moving to plasmids in a moving B. aphidicola ancestor. B. aphidicola became polyploidy at a given stage of its evolution and plasmid genes have been transferred to the main chromosome through several independent events.

Publication Types:

- Review

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3: Microbiol Mol Biol Rev. 2004 Dec;68(4):745-70. Related Articles, Links

Metabolic interdependence of obligate intracellular bacteria and their insect hosts.

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Mutualistic associations of obligate intracellular bacteria and insects have attracted much interest in the past few years due to the evolutionary consequences for their

genome structure. However, much less attention has been paid to the metabolic ramifications for these endosymbiotic microorganisms, which have to compete with but also to adapt to another metabolism--that of the host cell. This review attempts to provide insights into the complex physiological interactions and the evolution of metabolic pathways of several mutualistic bacteria of aphids, ants, and tsetse flies and their insect hosts.

Publication Types:

- Review

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4: Trends Microbiol. 2004 Jan;12(1):37-43. Related Articles, Links

ELSEVIER FULL-TEXT ARTICLE

Evolution of minimal-gene-sets in host-dependent bacteria.

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Several attempts have been made to identify the minimal set of genes that is required for life using computational approaches or studies of deletion mutants. These experiments resemble those already performed by nature; a few hundred million years ago an ancestor of Escherichia coli was domesticated by aphids, which resulted in the elimination of 70-75% of the original bacterial genome. Amazingly, the small genomes of these imprisoned bacteria are more stable than those of their free-living relatives. Minimal-gene-sets that have evolved naturally are largely species-specific, with the exception of a small set of core genes that are required for information processing. Comparative genomics of host-dependent bacteria have shown that minimal-gene-sets can persist in nature for tens of millions of years provided that the environment is rich in nutrients, that the host population size is large and that there is a strong host-level selection for bacterial gene functions.

Publication Types:

- Review

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5: Curr Opin Microbiol. 2003 Oct;6(5):512-8. Related Articles, Links

ELSEVIER FULL-TEXT ARTICLE

Tracing the evolution of gene loss in obligate bacterial symbionts.

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The gamma-proteobacterial symbionts of insects are a model group for comparative studies of genome reduction. The phylogenetic proximity of these reduced genomes to the larger genomes of well-studied free-living bacteria has enabled reconstructions of the process by which genes and DNA are lost. Three genome sequences are now available for Buchnera aphidicola. Analyses of Buchnera genomes in comparison with those of related enteric bacteria suggest that extensive changes including large deletions, repetitive element proliferation and chromosomal rearrangements occurred initially, followed by extreme stasis in gene order and slow decay of additional genes. This pattern appears to be characteristic of symbiont evolution.

Publication Types:

- Review

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6: Nat Rev Genet. 2002 Nov;3(11):850-61. Related Articles, Links

Genome evolution in bacterial endosymbionts of insects.

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Many insect species rely on intracellular bacterial symbionts for their viability and fecundity. Large-scale DNA-sequence analyses are revealing the forces that shape the evolution of these bacterial associates and the genetic basis of their specialization to an intracellular lifestyle. The full genome sequences of two obligate mutualists, Buchnera aphidicola of aphids and Wigglesworthia glossinidia of tsetse flies, reveal substantial gene loss and an integration of host and symbiont metabolic functions. Further genomic comparisons should reveal the generality of these features among bacterial mutualists and the extent to which they are shared with other intracellular bacteria, including obligate pathogens.

Publication Types:

- Review

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7: Genetica. 2002 May;115(1):1-12. Related Articles, Links

SpringerLink FULL-TEXT ARTICLE

Genome deterioration: loss of repeated sequences and accumulation of junk DNA.

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A global survey of microbial genomes reveals a correlation between genome size, repeat content and lifestyle. Free-living bacteria have large genomes with a high content of repeated sequences and self-propagating DNA, such as transposons and bacteriophages. In contrast, obligate intracellular bacteria have small genomes with a low content of repeated sequences and no or few genetic parasites. In extreme cases, such as in the 650 kb-genomes of aphid endosymbionts of the genus Buchnera all repeated sequences above 200bp have been eliminated. We speculate that the initial downsizing of the genomes of obligate symbionts and parasites occurred by homologous recombination at repeated genes, leading to the loss of large blocks of DNA as well as to the consumption of repeated sequences. Further sequence elimination in these small genomes seems primarily to result from the accumulation of short deletions within genic sequences. This process may lead to temporary increases in the genomic content of pseudogenes and 'junk' DNA. We discuss causes and long-term consequences of extreme genome size reductions in obligate intracellular bacteria.

Publication Types:

- Review

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8: Genome Biol. 2001;2(12):REVIEWS1032. Epub 2001 Nov 22. Related Articles, Links

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Genome interdependence in insect-bacterium symbioses.

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Symbioses between unicellular and multicellular organisms have contributed significantly to the evolution of life on Earth. As exemplified by several studies of bacterium-insect symbioses, modern genomic techniques are providing exciting new information about the molecular basis and the biological roles of these complex relationships, revealing for instance that symbionts have lost many genes for functions that are provided by the host, but that they can provide amino acids that the host cannot synthesize.

Publication Types:

- Review

PMID: 11790259 [PubMed - indexed for MEDLINE]

9: Acta Biochim Pol. 2001;48(2):367-81. Related Articles, Links

Reduction of bacterial genome size and expansion resulting from obligate intracellular lifestyle and adaptation to soil habitat.

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Prokaryotic organisms are exposed in the course of evolution to various impacts, resulting often in drastic changes of their genome size. Depending on circumstances, the same lineage may diverge into species having substantially reduced genomes, or such whose genomes have undergone considerable enlargement. Genome reduction is a consequence of obligate intracellular lifestyle rendering numerous genes expendable. Another consequence of intracellular lifestyle is reduction of effective population size and limited possibility of gene acquirement via lateral transfer. This causes a state of relaxed selection resulting in accumulation of mildly deleterious mutations that can not be corrected by recombination with the wild type copy. Thus, gene loss is usually irreversible. Additionally, constant environment of the eukaryotic cell renders that some bacterial genes involved in DNA repair are expandable. The loss of these genes is a probable cause of mutational bias resulting in a high A+T content. While causes of genome reduction are rather indisputable, those resulting in genome expansion seem to be less obvious. Presumably, the genome enlargement is an indirect consequence of adaptation to changing environmental conditions and requires the acquisition and integration of numerous genes. It seems that the need for a great number of capabilities is common among soil bacteria irrespective of their phylogenetic relationship. However, this would not be possible if soil bacteria lacked indigenous abilities to exchange and accumulate genetic information. The latter are considerably facilitated when housekeeping genes are physically separated from adaptive loci which are useful only in certain circumstances.

Publication Types:

- Review

PMID: 11732608 [PubMed - indexed for MEDLINE]

10: FEBS Lett. 2001 Jun 8;498(2-3):135-9. Related Articles, Links

ELSEVIER FULL-TEXT ARTICLE

Mutualists and parasites: how to paint yourself into a (metabolic) corner.

Tamas I, Klasson LM, Sandstrom JP, Andersson SG.

Department of Molecular Evolution, University of Uppsala, Norbyvagen 18C, S-752 36, Uppsala, Sweden.

Eukaryotes have developed an elaborate series of interactions with bacteria that enter their bodies and/or cells. Genome evolution of symbiotic and parasitic bacteria multiplying inside eukaryotic cells results in both convergent and divergent changes. The genome sequences of the symbiotic bacteria of aphids, Buchnera aphidicola, and the parasitic bacteria of body louse and humans, Rickettsia prowazekii, provide insights into these processes. Convergent genome characteristics include reduction in genome sizes and lowered G+C content values. Divergent evolution was recorded for amino acid and cell wall biosynthetic genes. The presence of pseudogenes in both genomes provides examples of recent gene inactivation events and offers clues to the process of genome deterioration and host-cell adaptation.

Publication Types:

- Review

PMID: 11412844 [PubMed - indexed for MEDLINE]

11: Annu Rev Microbiol. 1995;49:55-94. Related Articles, Links

ANNUAL REVIEWS

Genetics, physiology, and evolutionary relationships of the genus Buchnera: intracellular symbionts of aphids.

Baumann P, Baumann L, Lai CY, Rouhbakhsh D, Moran NA, Clark MA.

Microbiology Section, University of California, Davis 95616-8665, USA.

Evolutionary studies suggest that 200-250 million years ago an aphid ancestor was infected with a free-living eubacterium. The latter became established within aphid cells. Host and endosymbiont (genus Buchnera) became interdependent and unable to survive without each other. The growth of Buchnera became integrated with that of the aphids, which acquired the endosymbionts from their mothers before birth. Speciation of host lineages was paralleled by divergence of associated endosymbiont lineages, resulting in parallel evolution of Buchnera and aphids. Present day Buchnera retains many of the properties of its free-living ancestor, containing genes for proteins involved in DNA replication, transcription, and translation, as well as chaperonins and proteins involved in secretion, energy-yielding metabolism, and amino acid biosynthesis. Some of these processes are also observed in isolated endosymbiont cells. Genetic and physiological studies

indicate that Buchnera can synthesize methionine, cysteine, and tryptophan and supply these amino acids to the aphid host. In the case of some fast-growing species of aphids, the overproduction of tryptophan by Buchnera involves plasmid-amplification of the gene coding for anthranilate synthase, the first enzyme of the tryptophan biosynthetic pathway. These recent studies provide a beginning in our understanding of Buchnera and its role in the endosymbiosis with aphids.

Publication Types:

- Review

PMID: 8561471 [PubMed - indexed for MEDLINE]

Schedule of Experiments:

Date	Period	
Jan. 22	**1**	A. Instruction in use of the laboratory B. Preparation of Winogradsky columns
Jan. 29	**2**	Aseptic technique with the intestinal bacterium *Escherichia coli* (*E. coli* K12 laboratory strain)
Feb. 5	**3**	A. Measurements of light scattering and cell numbers for the intestinal bacterium *Escherichia coli* B. Analysis of results from the experiment of period 2
Feb. 12	**4**	Growth of *E. coli* and determination of its doubling times on enriched and minimal media
Feb. 19		**Presidents' Day, no lab.**
Feb. 26	**5**	Principles of light microscopy
Mar. 5	**6**	Use of stains for light microscopy
Mar. 12	**7**	Strain construction using phage P1-mediated transduction
Mar. 19	**8**	A. Assay of the activity of β-galactosidase, the product of the *lacZ* gene of *E. coli* B. Purification of lac^+ and trp^+ transductants: use of indicator plates
Mar. 26		**Spring break, no lab.**
Apr. 2	**9**	A. Induction of the lactose (*lac*) operon of *E. coli* and determination of the differential rate of synthesis of β-galactosidase, the product of the *lacZ* gene B. Growth of phage P1 / Analysis of transductants
Apr. 9	**10**	Assay of β-galactosidase in the cells from Period 9 (half of students in class) / Analysis of transductants (other half of students)
Apr. 16	**11**	Assay of β-galactosidase in the cells from Period 9 (second half of students in class) / Analysis of transductants (first half of students)
Apr. 23	**12**	Composting field trip (Oakland Compost Demonstration Garden)
Apr. 30	**13**	*E. coli* DNA microarrays and databases; PubMed
May 7	**14**	Conclusion/cleanup

*** We will meet on all Wednesday evenings between Jan. 17 and May 2 except Mar. 28 (Spring break).